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3053-Pos Board B100

Changes in the Secondary and Tertiary Structures of Secreted Phospholipase A₂ upon Activation

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Activation of human pancreatic phospholipase A₂ (PLA₂) in the presence of DPPC/DPPG (7:3) vesicles was induced by a temperature shift from 4 to 38 °C. PLA₂ activity was monitored by changes in fluorescence of bis-Pyrene-PC (2.5 mol % in the membranes), while simultaneous far- and near-UV circular dichroism (CD) spectra identified changes in the secondary and tertiary structures of the protein in real time. The 4-to-38 °C temperature shift caused dramatic changes in both bis-Pyrene-PC fluorescence and the protein CD spectra. The monomer fluorescence signal of bis-Pyrene-PC rapidly increased and the excimer signal decreased, demonstrating PLA₂ activation. Drastic weakening in the α -helical CD signal of the protein, i.e., a 20% decrease in the $n\text{-}\pi^*$ transition intensity at 222 nm, was detected upon enzyme activation. The α -helical signal exhibited a significantly smaller change upon a similar temperature shift under non-catalytic conditions (1 mM EGTA), while little changes were detected in the absence of lipid. Strong changes in the tertiary structure during PLA₂ activation were also identified. Initially, at 4 °C, the near-UV CD spectra showed a weak negative band around 280 nm. Upon a shift to 38 °C, strong positive CD bands rapidly developed around 250 and 280 nm, implying significant changes in the conformation and/or the micro-environment of Tyr and Trp side chains of PLA₂, possibly accompanied with a global tertiary structure perturbation associated with deformation of the abundant disulfide bonds in the protein. These experiments provide new information on the structure-function relationship of PLA₂ by near-simultaneous measurements of PLA₂ activity and its secondary and tertiary structures.

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Global Fitting and Kinetic Modeling of the Drug Transport Cycle of Human P-glycoprotein

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Human P-glycoprotein (ABCB1) is important in pharmacokinetics, tissue distribution, oral bioavailability and disposition of therapeutic compounds. Over-expression of P-glycoprotein can lead to adverse clinical effects such as the failure of cancer chemotherapy by the induction of multidrug resistance. Thus a detailed understanding of P-glycoprotein's drug transport cycle is a prerequisite to modulating its various activities. Here we present a detailed kinetic and mechanistic model of drug transport by P-glycoprotein. This was achieved by global fitting of time-based progress curves of the transport of spin-labeled verapamil as a function of ATP concentrations and measurements of the force-flux relationships observed during drug transport. By measuring the ATPase activity simultaneously with the transport velocity in proteoliposomes containing purified P-glycoprotein we established a limiting stoichiometry of one ATP hydrolyzed per spin-labeled verapamil molecule transported. Next, using standard chemical kinetic rate laws, we compared different proposed models of drug transport by trying to globally fit available experimental data numerically to coupled differential equations generated by each competing model. Our original seed values and constraints were generated from measurements of K_M , V_{max} , and K_i values for all reacting ligands during the steady state transport cycle together with knowledge of the overall thermodynamics of the transport cycle. We achieved a unique global fit of the progress curves of spin-labeled verapamil transport as a function of ATP concentration. A single internally consistent set of rate constants was shown to account for the data. Additionally, these same rate constants also accounted for the experimentally observed force-flux relationships when utilized in our "reaction power stroke" model. In contrast we were not able to fit the data as uniquely and as effectively with other competing drug transport models. Supported by NIH grant GM52502.

3055-Pos Board B102

Agonist and Antagonist Interactions in Beta Adrenergic Receptors

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G-protein coupled receptors (GPCRs) are a large family of integral membrane proteins involved in signal transduction pathways, making them ap-

pealing drug targets for a wide spectrum of diseases. The recently solved X-ray structures of beta1 (B1AR) and beta2 (B2AR) adrenergic receptors bound to inverse agonists/antagonists open up a large field of potential investigations to understand the binding modes and mechanisms of activation of GPCRs.

To investigate their structural and dynamic properties under pseudo in vivo conditions, we performed extensive molecular dynamics simulations (in an explicit membrane) of adrenergic receptors in complex with partial inverse agonists and agonists as well as in their apoform. To this end, we applied MD-based enhanced sampling techniques (steered MD, metadynamics, ...) to describe ligand binding and to elucidate the process of ligand entrance and release.

In this contribution, we rationalize the differences in binding mode between B1AR and B2AR for both agonists and antagonists (focusing on a limited set of key residues surrounding the binding pocket that are different between B1AR and B2AR). We also discuss main structural changes upon agonist/antagonist binding also in comparison with the most thoroughly studied GPCR, rhodopsin.

3056-Pos Board B103

Expression and functional characterization of Metabotropic Glutamate Receptor Type 6 (mGluR6) in detergent micelles

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Metabotropic glutamate receptors (mGluRs) are G protein coupled receptors which are implicated in different brain functions and dysfunctions including learning, memory, pain perception, neurodegeneration, schizophrenia and addiction. Metabotropic glutamate receptor type 6 (mGluR6) is a subtype of mGluRs which is exclusively expressed in ON-bipolar cells and is involved in night vision. Recent genome-wide association studies have discovered involvement of mGluR6 in heroin addiction. mGluR6 is emerging as a new drug target but structure-function relationships of this receptor and in general of mGluRs are poorly understood. These receptors have very low expression levels in their native cells and are only active in a membrane environment. The major problem in studying these receptors with biophysical approaches is in obtaining sufficient quantities of functional protein. To overcome this, we have constructed a tetracycline inducible mammalian stable cell line expressing full length human mGluR6. We optimized the detergent and buffer conditions required for mGluR6 purification. We are developing a reliable, quantitative *in vitro* assay for verifying the function of purified mGluR6 in different detergents. We have been successful in developing a medium scale expression and purification system of mGluR6. Preliminary data suggests that this receptor responds to a number of ligands relevant to its implicated role in vision and addiction with changes in activity and structure as evidenced by G protein binding and cysteine accessibility measurements, respectively.

3057-Pos Board B104

Calcium Enhances The Proteolytic Activity Of BACE: An In Vitro Biophysical And Biochemical Characterization Of The BACE-Calcium Interaction

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BACE is a novel type I transmembrane aspartyl protease that has been implicated in the pathogenesis of Alzheimer's disease. Cleavage of the amyloid precursor protein by the beta-secretase, BACE, is the first step in the production of the amyloid-beta peptide and is a prime target for therapeutic intervention. Using circular dichroism, we provide evidence that show differences in stability between active (pH 4.8) and inactive (pH 7.2) BACE. Active BACE ($T_m \sim 51^\circ\text{C}$) is comparably much less stable than the inactive form ($T_m \sim 84^\circ\text{C}$). In this study we have also examined Ca^{2+} binding to BACE, the effect of this binding on the secondary and tertiary structural characteristics of BACE, and the influence of this binding on the specific activity of the purified protein. Initially, we used isothermal titration calorimetry to characterize the Ca^{2+} -BACE interaction. Our results suggest that there is a high affinity of binding ($K = 2.0 \times 10^5 \text{ M}^{-1}$) between Ca^{2+} and BACE and that the binding process was exothermic (-3.5 kcal/mol). Circular dichroism and endogenous tryptophan fluorescence measurements demonstrated that the secondary and tertiary structure, respectively, is sensitive to increasing concentrations of Ca^{2+} . We also could demonstrate that low concentrations of Ca^{2+} (μM) significantly increased the proteolytic activity of BACE. Collectively, these results define a role for Ca^{2+} in both modulating the structure and proteolytic activity of BACE and suggest